

Application Note

Wyatt Technology Corporation

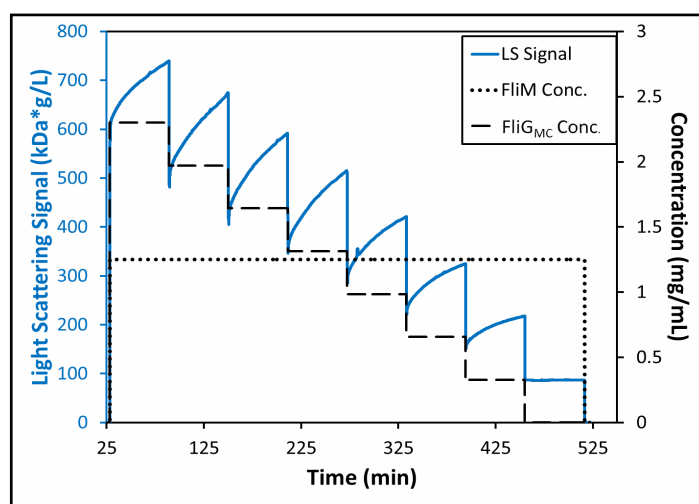
Revealing the Affinity of Individual and Combined FliG Domains for FliM in the Bacterial Flagellar Motor Complex

Summary

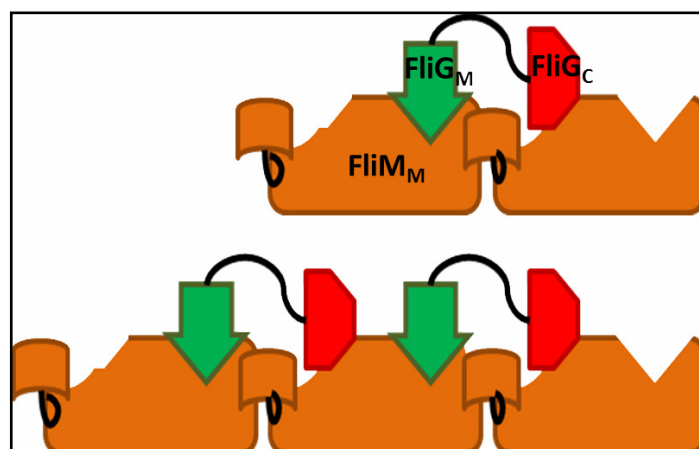
Complex interactions between proteins modulate the rotational direction of bacterial flagella. In particular, the middle and C-terminal domains of FliG (FliG_M and FliG_C, respectively) bind two different sites on the binding partner FliM, as part of the flagellar motor switch. We extend previous nuclear magnetic resonance (NMR) studies¹ of the interactions between FliG domains with FliM via composition-gradient multi-angle static light scattering (CG-MALS) to confirm specific binding, quantify affinities, and identify the stoichiometries of complexes formed.

For each FliG domain (FliG_M and FliG_C) interacting with FliM, separate automated composition gradients were created using a Calypso[®] II connected to a DAWN[®] HELEOS[®] MALS detector. A composition gradient of the multi-domain FliG_{MC} protein interacting with FliM was also performed. Using CG-MALS, we found FliG_M to have a strong interaction ($K_D = 6.6 \mu\text{M}$) and FliG_C to have a weak interaction ($K_D = 580 \mu\text{M}$) with FliM. Surprisingly, we found that the multidomain proteins assembled into complexes larger than 1:1 in a dramatically slower reaction that did not reach equilibrium after an hour.

CG-MALS provided insights into a complicated protein-protein interaction not possible with NMR or traditional techniques. The nearly 100-fold greater binding affinity of FliG_M over FliG_C for FliM supports a mechanism for changing the rotational direction of the flagellar motor in which FliG_C is displaced while FliG_M remains bound. The slow assembly of the multi-domain proteins into higher-order complexes uniquely captured by CG-MALS provides direction for future studies and may help determine the mechanism of flagellar motor switch self-assembly.



LS data for the interaction of multidomain FliG mixed with a constant concentration of FliM. Unexpectedly, this reaction shows slow, time-dependent association into complexes larger than 1:1.



Intermediate assembly states of the association of multidomain FliG and FliM derived from a quasi-equilibrium analysis. These intermediate species would most likely have continued forming longer FliM-FliG chains if the reaction had been allowed to continue.



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I. Introduction

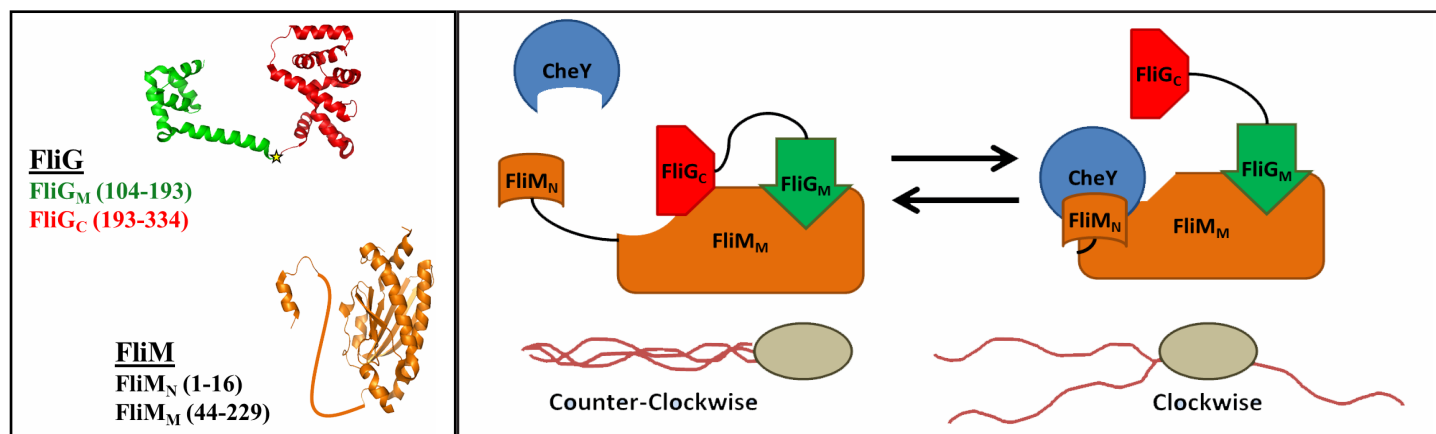


Figure 1: Interactions between flagellar motor proteins modulate the rotational direction of the bacterial flagella. Left: Flagellar motor proteins FliG and FliM³. Right: FliG binds two different sites on FliM as part of a proposed flagellar motor switch in which phosphorylated CheY displaces FliG_C. FliG_C's interaction with the stationary components of the motor generates torque and changes the rotational direction from counter-clockwise to clockwise (adapted from [1]).

A rotary motor composed of proteins organized into stacks of rings drives bacterial flagella.² The C-ring portion of the motor contains a group of proteins that form a directional switch. Complex interactions between the proteins in this ring and signaling proteins enable the motor's rotation to change from counter-clockwise to clockwise. Of specific interest are the middle and C-terminal domains of FliG (FliG_M and FliG_C, respectively) which bind two different sites on FliM (Figure 1).^{1,2} Previous studies of the interactions between FliG domains with FliM suggest a switch mechanism in which phosphorylated CheY displaces FliG_C while FliG_M remains bound.¹ The affinity of the individual domains of FliG (FliG_M and FliG_C) for FliM has been previously estimated by NMR and SEC-MALS. However, measurement of the interaction of the multidomain protein FliG with FliM by traditional methods is complicated by *in vitro* oligomerization of these proteins. We extend these studies via composition-gradient multi-angle static light scattering (CG-MALS) to confirm specific binding, measure equilibrium dissociation constants (K_D), and identify the stoichiometries of the complexes formed

prepared and delivered different compositions of protein and buffer to a Wyatt DAWN HELEOS MALS detector (Figure 2). Polycarbonate (Millipore) filter membranes with 0.1- μ m pore size were installed in the Calypso for sample and buffer filtration.

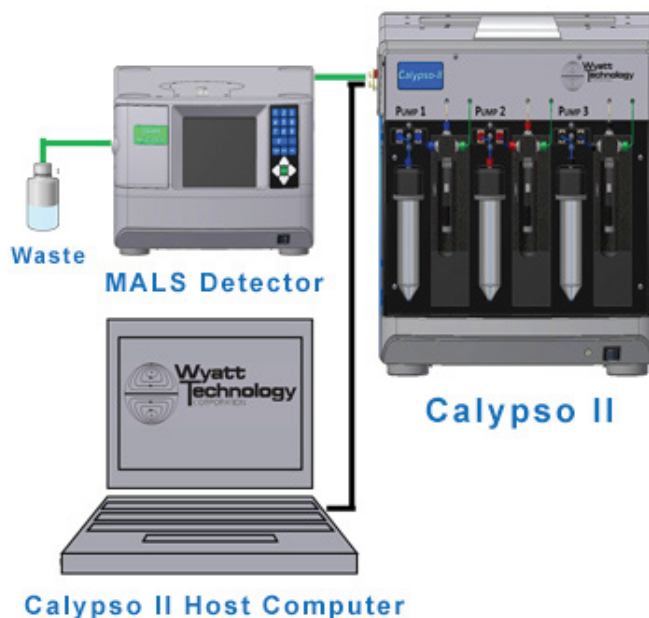


Figure 2: Calypso system hardware setup with inline DAWN HELEOS MALS detector.

II. Materials and Methods

Reagents and Instrumentation

FliG and FliM samples were kindly provided by Prof. Frederick Dahlquist at the University of California, Santa Barbara. All samples were prepared in Tris buffer filtered to 0.1 μ m (10 mM Tris, 100 mM NaCl, 1 mM EDTA, pH 7.5). After dilution to the appropriate concentration, samples were filtered to 0.02 μ m using Anotop syringe filters (Whatman) and the concentration determined by UV.

CG-MALS experiments were performed with a Calypso II composition-gradient system (Wyatt Technology) that



III. Results and Discussion

Determination of Equilibrium Dissociation Constant and Stoichiometry

Three sets of composition gradients were performed to quantify the interaction between FliM and three FliG constructs: the middle domain of FliG (FliG_M), the C-terminal domain of FliG (FliG_C), and the multidomain FliG_{MC}. Each experiment consisted of a dual-component “crossover” composition gradient to assess the hetero-association behavior and, if enough sample was available, single-component concentration gradients to quantify any self-association. Data collection and analysis of equilibrium association constants were performed using Calypso software. For each gradient, protein solution at the appropriate concentration was injected into the MALS detector. The flow was then stopped to allow the solution to come to equilibrium within the MALS flow cell. Specifics for each experiment are given below:

Stock Solution Concentration (mg/mL)			
FliM Binding Partner	FliM	Binding Partner	Stop-Flow Time (s)
FliG _M	0.23	0.15	500
FliG _C	4.86	4.44	500
FliG _{MC}	1.25*	2.3	3600

*The concentration of FliM was held constant throughout this hetero-association gradient, as shown in Figure 7.

Table 1: The stock solution concentration and the stop-flow times used for the three sets of composition gradient experiments.

Strong Interaction between FliM and FliG_M

Curvature in the light scattering (LS) signal in the “crossover” hetero-association gradients indicates interaction (Figure 3). Neither FliM nor FliG_M were found to self-associate under these conditions. For FliG_M interacting with FliM, the LS data from each injection were fit to a model (Figure 4) that included free FliM monomer, free FliG_M monomer, and a 1:1 complex (FliM)(FliG_M). FliG_M was found to interact with FliM with a binding affinity $K_D = 6.6 \mu\text{M}$. A previous analysis based on concentration and SEC analysis estimated a K_D for this interaction of 1-10 μM .

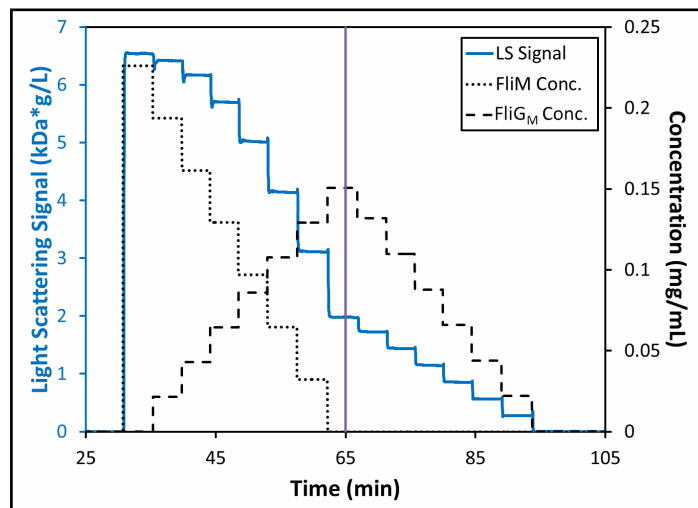


Figure 3: LS and concentration data for the interaction of FliM and FliG_M. The hetero-association “crossover” gradient occurs prior to 65 minutes, while the self-association of FliG_M is measured by the single-concentration gradient after 65 minutes.

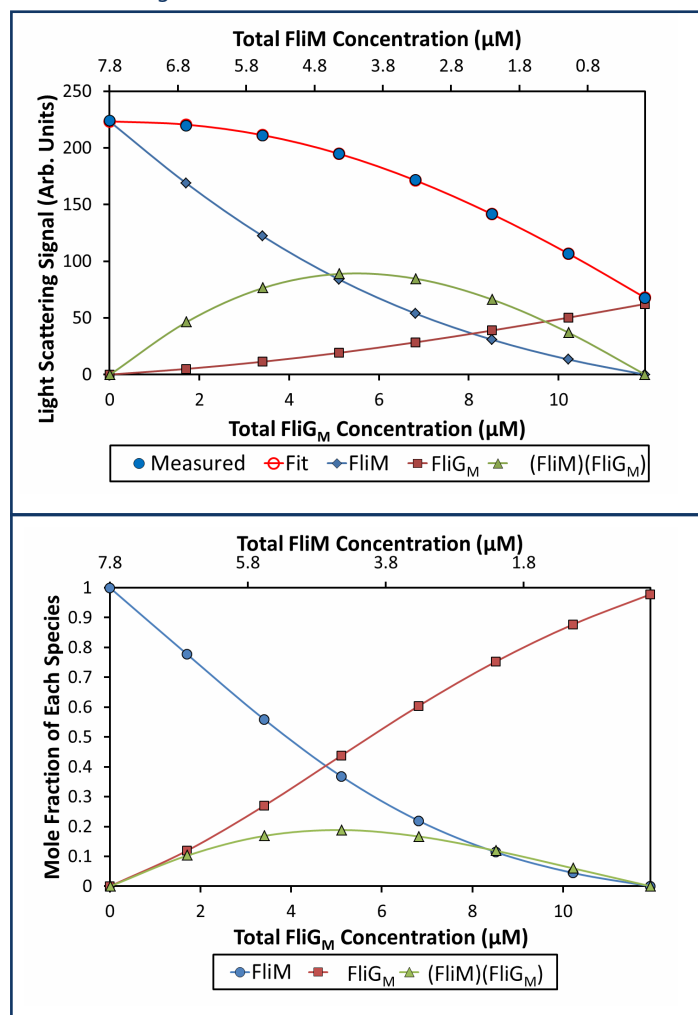


Figure 4: Top: LS data from the crossover region were fit to a model that included free FliM monomer, free FliG_M monomer, and 1:1 complex (FliM)(FliG_M). Bottom: Equilibrium concentrations of complex and monomers.



III. Results and Discussion (Continued)

Weak Interaction between FliM and FliG_C

The LS data (Figure 5) for FliM interacting with FliG_C were fit to a model (Figure 6) that included free FliM monomer, free FliG_C monomer, and a 1:1 complex (FliM)(FliG_C). The binding affinity of $K_D = 580 \mu\text{M}$ determined for FliG_C with FliM is in good agreement with the $K_D \sim 200 \mu\text{M}$ estimated by NMR.¹ Previous NMR and SEC-MALS studies indicated that domains of FliG differed in their affinity for FliM by at least an order of magnitude. CG-MALS determined the affinity of FliG_C for FliM to be 100-fold lower than that measured for FliG_M. The weaker interaction of FliG_C suggests that the phosphorylated CheY signaling protein ($K_D = 0.04 \mu\text{M}$)⁴ will easily replace it on FliM. On the other hand, FliG_M binds with higher affinity and is predicted to stay bound, holding the FliG protein in contact with FliM during motor switching.

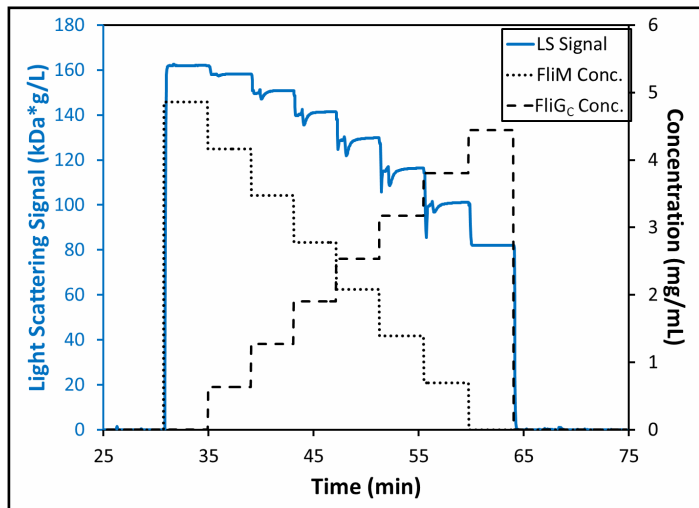
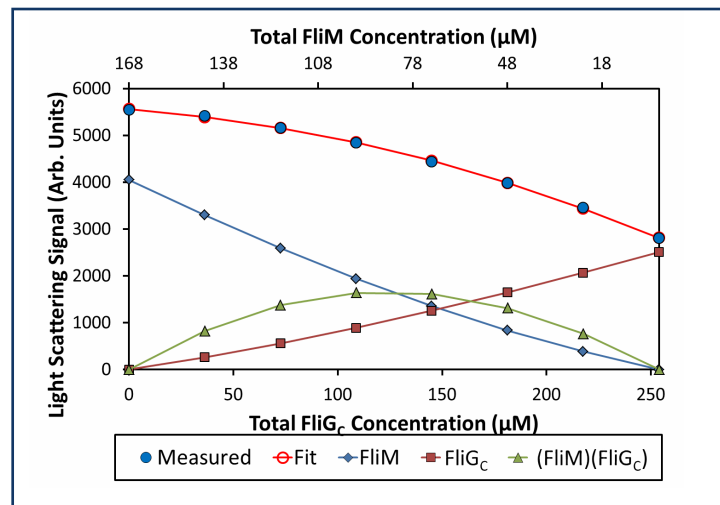


Figure 5: LS and concentration data for the interaction of FliM and FliG_C.



Complex Association of FliM with Multi-domain FliG

We assumed that multi-domain FliG would bind FliM with the same affinity as FliG_M or higher. However, the association between the multi-domain FliG protein and FliM is qualitatively distinct from single-domain interaction. The LS data exhibited slow, concentration-dependent association kinetics, with equilibrium not reached within 1 h (Figure 7). This is dramatically slower than individual FliG_M or FliG_C domain binding to FliM, both of which reached equilibrium within 20 s.

The kinetics of the multi-domain analysis were analyzed for relaxation time (τ) and amplitude (A) by fitting the data to the first-order exponential: $[R(t)/K] = (R/K)_{\infty} + Ae^{-t/\tau}$ (Figure 7). The equilibrium LS signal was extrapolated from the data for each injection and converted to an apparent weight average molar mass, M_w^{app} . The calculated M_w for FliM (70.2 kDa) was significantly higher than the predicted mass (28.9 kDa). In addition, an apparent molecular weight of 41 kDa was calculated for multidomain FliG from the pre-load light scattering signal which was significantly higher than the predicted mass of 27.5 kDa. This mismatch may have resulted from self-interactions, uncertainty in the stock concentration, or both.

The M_w^{app} was calculated for each injection at three different time points: $t \sim 0$, $t \sim 30$ min, and $t \sim 55$ min (Figure 8). It appears to be asymptotically approaching a value of ~ 230 kDa. This value is more than the molecular weight for a fully associated 1:1 complex (assuming the measured monomer molecular weights of 70 and 41 kDa), indicating the formation of larger complexes.

A quasi-equilibrium analysis was performed using the 1-h data points and the extrapolated data. The best fit included species with 2:1 and 3:2 FliM:FliG stoichiometries with an average per-site affinity of $10 \mu\text{M}$. The slow increase

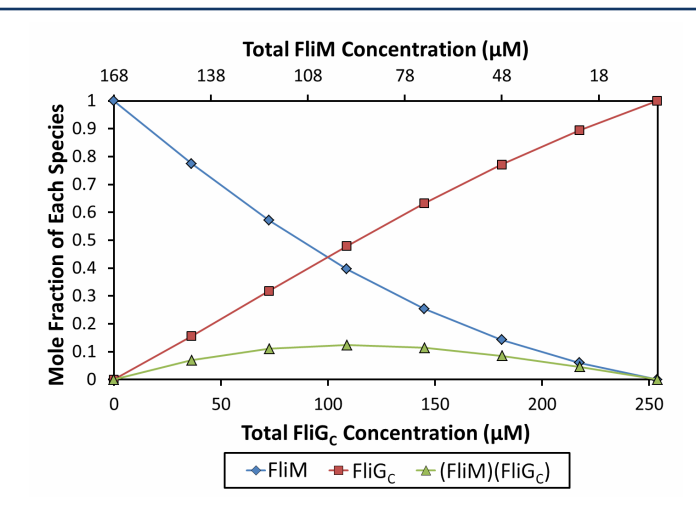


Figure 6: LS data from the crossover region were fit to a model that included free FliM monomer, free FliG_C monomer, and the (FliM)(FliG_C) complex. Bottom: Equilibrium concentrations of complex and monomers.



III. Results and Discussion (Continued)

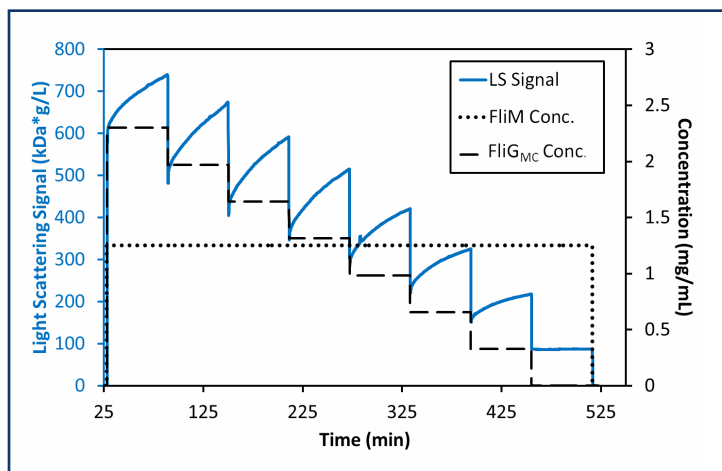


Figure 7: Multidomain FliG association with FliM. Left: The LS data exhibited slow concentration-dependent association kinetics, with equilibrium not reached within 1 h. Right: Kinetics parameters and equilibrium M_w^{app} were determined for each injection by fitting the data to first-order kinetics. Injection #8 consisted of pure FliM, which displayed no association in the absence of FliG.

of the LS signal until the next injection suggests that these intermediate species would continue forming longer FliM-FliG chains until the protein supply was exhausted if the reaction had been allowed to continue.

The observed *in vitro* kinetics are not likely to be relevant to *in vivo* motor assembly which could be assisted by chaperones, biological machinery, or scaffolding. However, it is interesting that C-ring assembly initiates even without these supporting factors. Understanding the slow association of FliM and FliG into large complexes may help determine the mechanism this self-assembly.

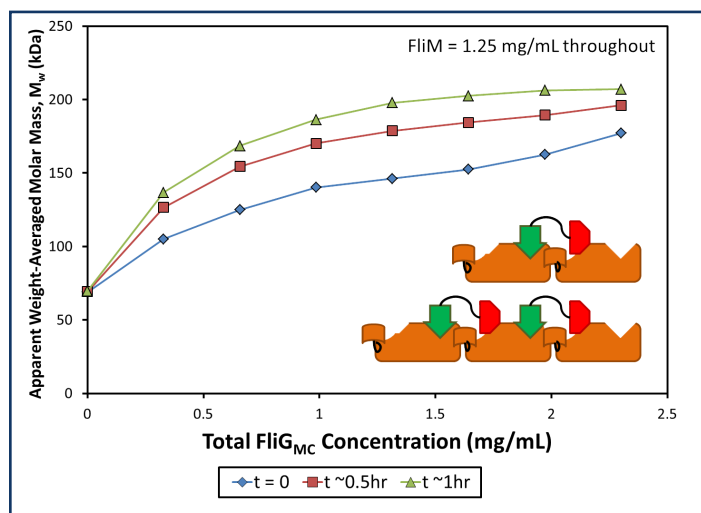


Figure 8: The calculated M_w^{app} appears to be reaching an equilibrium with FliM:FliG stoichiometry >1:1. Inset: Intermediate assembly states derived from a quasi-equilibrium analysis.

References

- (1.) Dyer C.M., et al. J. Mol. Biol. (2009) 388, 71.
- (2.) Blair, D.F. J. Bacteriol. (2006) 188, 7033.
- (3.) FliG PDB ID: 1LKV; FliM PDB ID: 2HP7.
- (4.) Park, S., et al. Proc. Natl. Acad. Sci. (2006) 103, 11886.

Conclusion

CG-MALS provided new insights into the binding of FliG to FliM, two key proteins involved in the bacterial flagellar motor switching mechanism. We found a 100-fold difference in binding affinity between FliG_M ($K_D = 6.6 \mu\text{M}$) and FliG_C ($K_D = 580 \mu\text{M}$) for FliM. This large difference in affinity supports a scheme for changing the rotational direction of the flagellar motor wherein FliG_C is displaced while FliG_M remains bound.

Initial expectations for the interaction of the multidomain FliG protein with FliG_M suggested formation of a 1:1 complex that would exhibit similar kinetics to the single-domain binding experiments. Instead, the multidomain proteins assembled into complexes larger than 1:1 in a reaction that did not reach equilibrium on the time scale of the experiment.

Surface techniques such as surface plasmon resonance (SPR), which requires one binding partner to be immobilized on a surface, preclude true solution behavior wherein more than one molecule of each protein is required in the final complex. Most solution-based measurements, such as isothermal titration calorimetry (ITC), only determine stoichiometric ratios and so would not provide conclusive evidence for the 3:2 complex. Sedimentation equilibrium—which, like CG-MALS, measures molar mass—could in principle indicate the formation of large complexes but would not track real-time self-assembly. The ability of CG-MALS to comprehensively quantify the slow, time-dependent association of FliG and FliM into large complexes is uniquely powerful and may be applied to many other difficult biomolecular systems.

